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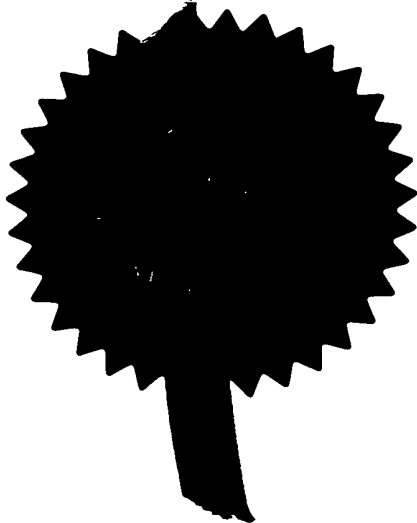
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KEG/RJL/40244 290CT98 E400542-5 D01631

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2. Patent application number
(The Patent Office will fill in this part)

27 OCT 1998

9823515.3

3. Full name, address and postcode of the or of each applicant (underline all surnames)

Ferring B.V.,
Marsstraat 9,
P. O. Box 3129,
2130 KC Hoofddorp,
THE NETHERLANDS

Patents ADP number (if you know it)

If the applicant is a corporate body, give the country/state of incorporation

7012081001

4. Title of the invention

Agent for the Treatment of Osteoporosis

5. Name of your agent (if you have one)

Reddie & Grose
16 Theobalds Road
LONDON
WC1X 8PL

"Address for service" in the United Kingdom to which all correspondence should be sent (Including the postcode)

91001

Patents ADP number (if you know it)

6. If you are declaring priority from one or more earlier patent applications, give the country and the date of filing of the or of each of these earlier applications and (if you know it) the or each application number

Country

Priority application
(If you know it)

Date of filing
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Number of earlier application

Date of filing
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8. Is a statement of inventorship and of right to grant of a patent required in support of this request? (Answer 'Yes' if:

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Continuation sheets of this form

Description 12
Claim(s) 02
Abstract
Drawing(s)

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Priority documents NO
Translations of priority documents NO
Statement of inventorship and right to grant of a patent (*Patents Form 7/77*) NO
Request for preliminary examination and search (*Patents Form 9/77*) YES
Request for substantive examination (*Patents Form 10/77*) NO
Any other documents (please specify) NO

11.

I/We request the grant of a patent on the basis of this application.

Signature

Date

Reddie & Grose

27 October 1998

12. Name and daytime telephone number of person to contact in the United Kingdom

K E GEERING - Reddie & Grose
0171-242 0901

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5021-18

AGENT FOR THE TREATMENT OF OSTEOPOROSIS

FIELD OF INVENTION

The present invention relates to pharmaceutical compositions for the treatment of osteoporosis and other disorders of bone metabolism.

BACKGROUND TO THE INVENTION

i) Osteoporosis

Osteoporosis is a disease which affects many elderly people. Post-menopausal osteoporosis associated with oestrogen deficiency is the most common bone disease in the western world (Melton *J. Bone Miner. Res.* 10 174-177, 1995). The basic mechanism underlying osteoporosis is when bone resorption exceeds bone formation resulting in net bone loss and an increase in the incidence of fractures. The dynamic processes that control bone morphogenesis take place in bone and bone marrow. Bone is a hard calcified connective tissue consisting of osteoprogenitor cells, osteoblasts, osteocytes and osteoclasts. Bone marrow is composed of haematopoietic stem cells and mesenchymal cells. These cells include osteoprogenitors, chondrocytes, adipocytes, fibroblasts and reticular cells. Bone metabolism and remodelling is dependent on a balance between two processes; bone resorption is dependent on osteoclast activity and bone formation is dependent on osteoblast activity. Osteoblast precursors are fibroblast-like, proliferating progenitor cells, which differentiate into osteoblasts that synthesise and deposit matrix to become new bone. Osteoclasts are a tissue specific subtype of macrophages responsible for resorption of bone. Mononuclear phagocytes from bone marrow fuse to form the large multinucleated osteoclasts. Osteoclastogenesis is a complex phenomenon that is facilitated by the interaction of bone marrow stromal cells with haematopoietic osteoclast precursors (Suda *et al.*, Endocr. Rev. 13 66-80, 1992)

A number of factors regulate the proliferation and differentiation of osteoblasts and osteoclasts including cell-cell interactions, cytokines, growth factors, steroid hormones and neurohormones. These factors act via endocrine, autocrine and paracrine mechanisms. Parathyroid hormone (PTH) is a key systemic regulator of calcium and bone metabolism acting principally by regulating bone resorption. There is evidence to suggest that PTH, vitamin D and calcitonin actions may be mediated by local factors found in bone (Oursler *et al.*, Endocrinol. 129 3313-3320, 1990). A number of growth factors have been implicated such as insulin-like growth factors (IGF-I and IGF-II), fibroblast growth factors (bFGF and aFGF) (Baylink *et al.*, J. Bone Mineral Res. 8 (Supp. 2) S565-S572, 1993). Identification of locally produced agents that influence the process of bone formation and bone resorption may provide alternatives to current

treatments for osteoporosis which include salmon calcitonin and hormone (oestrogen and progesterone) replacement therapy (Lindsay *et al.*, Lancet **341** 801-805, 1993)

ii) **GnRH-II**

Studies on the physiology of the hypothalamic-pituitary-gonadal axis have resulted in the recognition of gonadotropin releasing hormone (GnRH, otherwise known as luteinizing hormone releasing hormone, LHRH) as a key regulatory hormone. GnRH is released by the hypothalamus and acts on the pituitary to stimulate the release of luteinizing hormone (LH) and follicle-stimulating hormone (FSH). More recently, a peptide related to GnRH has been identified (White *et al.*, Proc. Natl. Acad. Sci. USA **95** 305-309, 1998). This peptide has been called GnRH-II. The sequences of the two peptides are compared below.

GnRH pyroGlu-His-Trp-Ser-Tyr-Gly-Leu-Arg-Pro-Gly-NH₂

GnRH-II pyroGlu-His-Trp-Ser-His-Gly-Trp-Tyr-Pro-Gly-NH₂

No clear evidence for a physiological role for GnRH-II has so far been presented.

SUMMARY OF THE INVENTION

We have now found that GnRH-II is capable of modulating the differentiation of bone precursor cells and inducing the expansion of osteoblast populations. Accordingly, it is an object of the present invention to provide a pharmaceutical composition for the treatment of osteoporosis, which composition is characterised by the inclusion of GnRH-II or an analogue thereof. More specifically, the composition includes a peptide according to the sequence

pyroGlu-His-Trp-Ser-Xaa¹-Gly-Xaa²-Xaa³-Pro-Gly-NH₂

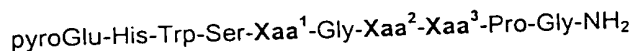
wherein Xaa¹ is His or Tyr,
 Xaa² is Trp or Leu, and
 Xaa³ is Tyr or Arg,

provided that when Xaa¹ is Tyr and Xaa² is Leu, then Xaa³ is not Arg. The invention further comprises a method for the preparation of such a composition, wherein the active peptide is combined with pharmaceutically acceptable excipients, and a method of treatment of osteoporosis, which is the administration to an individual in need of such treatment a therapeutically effective amount of the composition.

DESCRIPTION OF THE INVENTION

As used herein, abbreviations referring to amino acids have their conventional meanings and indicate the natural L-isomer (except for the achiral amino acid glycine).

In a first embodiment, the invention as disclosed herein comprises a pharmaceutical composition for increasing bone mass or bone density, or for accelerating bone growth or repair. Preferably, the invention as disclosed herein comprises a pharmaceutical composition for the treatment of osteoporosis. The composition is characterised in that it includes as an active principal a peptide according to the sequence

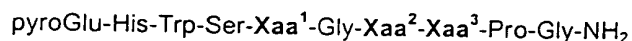


wherein Xaa^1 is either His or Tyr, Xaa^2 is either Trp or Leu, and Xaa^3 is either Tyr or Arg, provided that when Xaa^1 is Tyr and Xaa^2 is Leu, then Xaa^3 is not Arg. Preferably, Xaa^1 is His, Xaa^2 is Trp, and Xaa^3 is Tyr. It will be recognised that such a peptide can form salts with acids (for example, acetic acid, trifluoroacetic acid, benzoic acid, hydrochloric acid, phosphoric acid and the like). To the extent that such salts are formed with pharmaceutically acceptable acids, they are included within the scope of the invention.

It will be understood by those familiar with the art that the composition will also comprise one or more excipients, such as diluents, carriers, preservatives, and the like. The composition may be a solid or liquid formulation, and will be appropriate for the intended route of administration. For example, the composition might be formulated as a tablet or capsule for oral administration, or as a suppository for rectal or vaginal use. Equally, the composition might be formulated as a solution, for example in isotonic saline, for intravenous administration. The composition may also be presented as a kit for formulation immediately prior to administration, for example as a freeze-dried powder in a sealed ampoule supplied with a second ampoule containing a solvent such as saline.

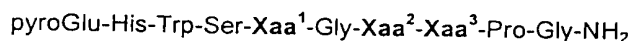
The composition may be used in clinical situations other than for the treatment of osteoporosis, where bone mass or bone density are reduced, or where bone growth or repair needs to be accelerated. Such situations include (but are not limited to) the treatment of osteogenesis imperfecta and osteomalacia, the prevention of bone loss when an individual is immobilised for an extended period, and the facilitation of healing following a fracture.

In a second embodiment, the invention disclosed herein comprises a method for the preparation of a pharmaceutical composition for the treatment of osteoporosis, which method comprises the mixing of a peptide according to the sequence



wherein Xaa^1 , Xaa^2 and Xaa^3 are as defined above, with one or more pharmaceutically acceptable excipients. In the context of this invention, mixing is taken to include the blending together of solids, the dissolution of one or more solids in a liquid, and the dispersion of a solid in a liquid. The method may also include such processes as freeze-drying and microencapsulation necessary to obtain the formulation to be presented. Such processes as are known in the art are included within the scope of the present invention.

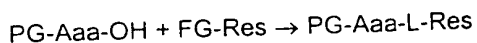
In a third embodiment, the invention as disclosed herein comprises a method for the treatment of an individual suffering from osteoporosis, or considered to be at risk of so suffering. This method of treatment comprises the administration to said individual of a therapeutically effective amount of a composition containing, as an active principal, a peptide according to the sequence



wherein Xaa^1 , Xaa^2 and Xaa^3 are as defined above. The method of treatment may comprise a single administration of the composition, but is more likely to comprise a course of repeated administrations. The frequency of the administrations may be from once per month to four times per day. The amount of active peptide in each dose will depend on the dosing schedule and the route of administration. Generally, it will be between one microgram (1 μg) and one hundred milligrams (100 mg). Doses in the range 1-100 μg are preferred when the composition is administered by intravenous injection. Doses in the range 10 μg - 10 mg are preferred when the composition is administered as slow-release depots. Doses in the range 1-100 mg are preferred when the composition is administered orally. Furthermore, high doses and frequent administration will be preferred when the desired outcome is to increase bone mass while lower doses and less frequent administration are more suited to the maintenance of bone mass.

The peptides that comprise the active agents of the compositions of the present invention can be prepared by the methods generally known in the art. For example, the peptides may be prepared by solid-phase synthesis. This involves the sequential addition of amino acid residues to a resin-bound intermediate according to the following strategy.

1. Formation of resin-bound first intermediate



Aaa = amino acid

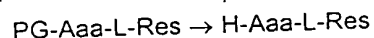
PG = protecting group

FG = functional group

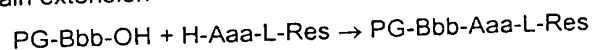
Res = polymeric resin

L = linker group (-O- or -NH-)

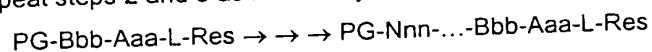
2. Deprotection



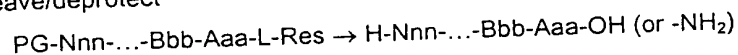
3. Chain extension



4. Repeat steps 2 and 3 as necessary



5. Cleave/deprotect



In step one, a-protected amino acid is reacted with a functionalised resin. The protecting group (PG) is most commonly *tert*-butyloxycarbonyl (Boc) or 9-fluorenylmethyloxycarbonyl (Fmoc). The functional group on the resin (FG) is commonly a chloroalkyl group, a hydroxyl group or an amine group. When FG is a chloroalkyl or hydroxyl group, the linker group (L) is an oxygen atom (-O-). When FG is an amine group, L is -NH-.

In step two, the protecting group (PG) is removed from the α -amino group. When PG is Boc, this can be accomplished by treating the resin with acids such as trifluoroacetic acid or hydrogen chloride in dichloromethane. When PG is Fmoc, the deprotection can be accomplished by treating the resin with bases such as piperidine.

In step three, the peptide chain is extended by one amino acid residue. A protected amino acid is coupled to the amine group liberated in step two. Many reagents are known in the art for achieving this conversion. One combination is dicyclohexylcarbodiimide (DCC) and hydroxybenzotriazole (HOBt). Generally, a base will also be necessary. Suitable bases include triethylamine and *N,N*-diisopropylethylamine. The solvent will generally be dichloromethane, dimethylformamide, or a mixture of these.

If the side chains of the amino acids (Aaa - Nnn) contain reactive groups (for example amino groups, carboxylic acid groups, hydroxyl groups) then these will need protecting. The protecting groups chosen for the side chains are generally those that are stable under the conditions required to remove the protecting group (PG) from the α -amino group. If PG is Fmoc, then the side chain protecting groups can conveniently be based on tert-butyl chemistry. On the other hand, if PG is Boc, then the side chain protecting groups can be based on fluorenylmethyl chemistry. Other protecting groups known in the art can also be used.

In step four, the deprotection/chain extension cycle is repeated until the desired peptide sequence has been constructed.

In step five, the completed peptide is liberated from the resin. Protecting groups are removed from the side chains either before or after the cleavage. When L is -NH-, the peptide liberated is in the form of the C-terminal amide. When L is -O-, the peptide liberated is often the C-terminal free acid and a second step is required to form the C-terminal amide.

The peptides may also be prepared by solution-phase synthesis, and this may be more convenient when large quantities of material are needed.

The above general description is further elaborated below in a number of examples. These are intended to illustrate certain aspects of the invention. They are not intended to be limiting in any way.

EXAMPLES

Example 1 - Synthesis of GnRH-II

1A. Preparation of resin-bound protected peptide.

pyroGlu-His(Bom)-Trp(CHO)-Ser(Bzl)-His(Bom)-Gly-Trp(CHO)-Tyr(Bzl)-His(Bom)-Pro-Gly-ORes

This peptide was prepared using standard solid-phase methods starting from Boc-Gly-esterified Merrifield resin (60 g, 1 mmol/g). The synthesis was performed in a manual synthesizer, with a total solvent and reagent volume of 300 mL for each operation. The standard deprotection/wash/coupling protocol is summarised in Table 1.

Table 1

Step	Reagent	Time (min)	Number of operations
Deprotection of Boc	HCl / DCM*	60	1
Washing	DCM	2 - 4	3
Neutralisation	10% DIPEA / DCM	4	2
Washing	DCM	2 - 4	1
Coupling	Activated ester	60 - 120**	1 - 2
Washing	DCM	2 - 4	3
* Gaseous hydrogen chloride was bubbled through a suspension of the resin in DCM			
** Completeness of reaction was determined by a negative ninhydrin test			

Benzotriazolyl esters were used as the activated esters throughout the synthesis. These were prepared from the corresponding protected amino acids by reaction with 1-hydroxybenzotriazole (1 eq.) and dicyclohexylcarbodiimide (1 eq.). The quantities used (in relation to the resin substitution capacity) are listed in Table 2.

Table 2

Cycle no.	Amino acid derivative	Molar excess
1	Boc-Pro-OH	1.8
2	Boc-Tyr(Bzl)-OH	1.8
3	Boc-Trp(CHO)-OH	1.8
4	Boc-Gly-OH	1.8
5	Boc-His(Bom)-OH	1.8
6	Boc-Ser(Bzl)-OH	2.0
7	Boc-Trp(CHO)-OH	2.0
8	Boc-His(Bom)-OH	2.0
9	pyroGlu-OH	2.0

Following the final coupling, the resin was washed with dichloromethane (3 × 3 L) and dried under reduced pressure at +40°C to constant weight.

Amino acid analysis: Consistent with proposed sequence

1B. Cleavage and deprotection

pyroGlu-His-Trp-Ser-His-Gly-Trp-Tyr-His-Pro-Gly-NH₂

The peptidoresin prepared in Example 1A was placed in a linen bag in a pressure vessel. The vessel was then charged with gaseous ammonia to a final pressure of 4 atm. After 72h the excess ammonia was vented and the resin was extracted with acetic acid (3×100mL) and ethanol (3×100mL). The combined extracts were degassed with nitrogen, 10% palladium-on-carbon was added, and the mixture was stirred under an atmosphere of hydrogen. When the reaction was complete (as judged by HPLC), the mixture was filtered and the filtrate was evaporated. The residue was purified by reverse-phase HPLC to give the title compound.

Example 2 - Analysis of the effects of GnRH-II and analogues on Osteogenic cell populations *in vitro*.

- (a) Human osteoblasts were isolated from cancerous bone from orthopaedic surgery (Nilsson *et al.*, 1995) according to standard procedures known in the art. The bone explants were minced into small bone chips and then washed extensively in Dulbecco's modified Eagle's medium (DMEM)/F12 (1:1 Gibco, Paisley, U.K). These osteoblast like cells, Murine osteoblastic MC3T3-E1 cells and human clonal osteosarcoma cell lines MG-63 (non-mineralising) and SaOS-2 (mineralising osteosarcoma) were cultured in DMEM:F12, 1:1 with the addition of 10% fetal calf serum (FCS, Gibco), fungizone (500mg/l), gentamycin sulphate (50mg/l), L-glutamine (2mM) and l-ascorbic acid (100mg/l) in a humidified CO₂ chamber at 37°C.
- (b) Human bone marrow stromal cells were isolated from bone fragments rinsed in phosphate-buffered saline. Bone marrow cells were collected and spun through a column of Ficoll Hypaque (Kimble *et al J. Clin. Invest.* 93 1959-1967, 1994). Cells at the interface were pelleted, counted and seeded into 75cm² flasks. The cells were incubated in a humidified CO₂ chamber at 37°C and the medium changed weekly. At confluence, the cells were harvested using trypsin EDTA and re-seeded in α -minimum essential medium (-MEM) supplemented with 10% fetal calf serum (FCS, Gibco), penicillin (100U/ml), streptomycin (100mg/ml), fungizone and L-glutamine (2mM).
- (c) All cells were serum-starved for 48h before addition of GnRH-I and GnRH-II. Cells were placed in DMEM without phenol red (in order to avoid oestrogen-like effects of phenol red)

containing 10% charcoal-stripped serum for 48 hours in 12 well plates. Dose dependent effects of GnRH-I and GnRH-II and analogues of the peptides were studied following the addition of peptides at final concentrations ranging from 10^{-9} to 10^{-6} M. 1mM dibutyl cAMP was used as a control. The cells were incubated for 24, 48 and 96h with the peptide being replaced every 24 hours.

- (d) To assess the effects of the peptides on cell proliferation, [3 H]thymidine was added at 1mCi/ml for an additional 24hours and [3 H]thymidine incorporation was determined. Radioisotope incorporation was determined using a scintillation counter and the results were calculated as cpm/mg of total protein.
- (e) Expression of osteoblastic differentiation markers was also determined (Tintut Y *et al.*, J Biol Chem 273 7547-53, 1998). Total RNA was isolated at several stages; before treatment, at 24, 48, 72 and 96 hours after addition of peptides. Type I procollagen, osteopontin and 28S RNA (used as an internal control) expression was determined by Northern blot analyses. Alkaline phosphatase, matrix GLA protein, osteoclastin and GAPDH (as an internal control) were determined by RT-PCR with specific primers designed for each gene.

The peptides of the invention caused significant effects at concentrations below 100 μ M.

Example 3 - Analysis of the effects of GnRH-II and analogues on Osteoclast populations *in vitro*.

- (a) Human clonal cell lines of osteoclast precursors (FLG 29.1) were used as an *in vitro* model of osteoclast differentiation (Gattei V *et al.*, Cell Growth Differ 7 753-63, 1996). In addition, co-cultures of FLG 29.1 and osteoblastic cells (Saos-2) were evaluated for migratory, adhesive, cytochemical, morphological, and biochemical changes. Dose dependent effects of GnRH-I and GnRH-II and analogues of the peptides were studied following addition at final concentrations ranging from 10^{-9} to 10^{-6} M to FLG 29.1 cultures and to co-cultures. Parathyroid hormone was added as a control. Potentiation (or inhibition) of the differentiation of the preosteoclasts (fusion into large multinucleated elements) and a number of other factors were measured (Orlandini *et al.*, Cell Tissue Res. 281 33-42, 1995). These included:

1. Positive staining for tartrate-resistant acid phosphatase in FLG 29.1 cells
2. A decrease of the alkaline phosphatase activity expressed by Saos-2 cells
3. The appearance of typical ultrastructural features of mature osteoclasts in FLG 29.1 cells
4. The release into the culture medium of granulocyte-macrophage colony stimulating factor.

5. To assess the effects the peptides on cell proliferation, [^3H]thymidine was added at 1mCi/ml for an additional 24hours and [^3H]thymidine incorporation was determined as described above.
- (b) Bone marrow cells removed from human bone fragments were cultured in the presence of 10nM 1,25-(OH) $_2$ vitamin D $_3$ for seven days to generate multinucleated osteoclasts using standard techniques known in the art (Takahashi *et al.*, Endocrinol 122 1473-1482, 1988). The culture medium (α -MEM) was removed and replaced by a fresh phenol red free medium supplemented with antibiotics and 10% charcoal-stripped heat-inactivated FCS containing GnRH-I, GnRH-II or analogues, and the cultures were maintained for a further 24 hours. Floating cells were harvested and osteoclasts stained for tartrate-resistant acid phosphatase (TRAP) expression, a marker of osteoclast differentiation (Hughes *et al.*, Nat. Med. 2 1132-1135, 1996)
1. Cells were incubated in 0.2M acetate buffer, pH 4.7-5.0, containing tartaric acid and 2% naphthol AS-BI phosphate (dissolved at 20mg/ml in ethylene glycol monomethyl ether) for 15min at 37°C. The cells were then transferred to a second solution consisting of the same buffer and concentration of tartaric acid with 0.1% pararosaniline chloride (hexazotised by mixing with an equal volume of 4% sodium nitrite for 5min at room temperature) for 10min at 37°C. This treatment causes a red cytoplasmic stain in cells expressing TRAP. Harris' hematoxylin was used as a nuclear counterstain.
 2. Apoptotic multinucleated osteoclasts were identified by strong expression of TRAP, larger size than accompanying viable TRAP-positive cells. Confirmation of apoptosis was carried out using acridine orange stain. Viable osteoclasts were counted after fixation in 95% ethanol and TRAP hematoxylin staining, and apoptotic osteoclasts were expressed as a percentage of the total number of multinucleated osteoclasts (viable and apoptotic) in each culture well.

The peptides of the invention caused significant effects at concentrations below 100 μM .

Example 4- Expression analysis of GnRH ligands in osteogenic and osteoclast cell populations

Total RNA was extracted from cells cultured as described above:

1. osteoblast like cells, isolated from cancerous bone
2. murine osteoblastic MC3T3-E1 cells
3. MG-63 (non-mineralising)
4. SaOS-2 (mineralising osteosarcoma)
5. human bone marrow stromal cells
6. human FLG 29.1 osteoclast precursor cells
7. multinucleated osteoclasts generated from bone marrow

Expression of GnRH-I and GnRH-II was determined by RT-PCR using PCR primers outlined in SEQ I.D. No 1-4. The integrity of the cDNA generated was determined by assessing the relative level of actin amplification.

SEQUENCE LISTING

INFORMATION FOR SEQ ID No 1.

- (i) Sequence Characteristics
 - (a) Length: 19 bases
 - (b) Type: nucleic acid
 - (c) Strandedness: single
 - (d) Topology: linear
- (ii) Molecule Type cDNA
- (iii) Sequence description: SEQ ID No.1
CTG CAG CTG CCT GAA GGA G

INFORMATION FOR SEQ ID No 2.

- (i) Sequence Characteristics
 - (a) Length: 19 bases
 - (b) Type: nucleic acid
 - (c) Strandedness: single
 - (d) Topology: linear
- (ii) Molecule Type cDNA
- (iii) Sequence description: SEQ ID No.2
GGG CGG GGC GGG GCT CTC G

INFORMATION FOR SEQ ID No 3.

- (i) Sequence Characteristics
 - (a) Length: 21 bases
 - (b) Type: nucleic acid
 - (c) Strandedness: single
 - (d) Topology: linear
- (ii) Molecule Type cDNA
- (iii) Sequence description: SEQ ID No.3
ATT CTA CTG ACT TGG TGC GTG

INFORMATION FOR SEQ ID No 4.

- (i) Sequence Characteristics
 - (a) Length: 21 bases
 - (b) Type: nucleic acid
 - (c) Strandedness: single
 - (d) Topology: linear
- (ii) Molecule Type cDNA
- (iii) Sequence description: SEQ ID No.4
GGA ATA TGT GCA ACT TGG TGT

CLAIMS

1. A pharmaceutical composition for the treatment of osteoporosis and other disorders of bone metabolism, or for accelerating bone growth or repair, which composition is characterised by the inclusion of a peptide

pyroGlu-His-Trp-Ser-Xaa¹-Gly-Xaa²-Xaa³-Pro-Gly-NH₂

or a salt thereof

wherein Xaa¹ is His or Tyr,
Xaa² is Trp or Leu, and
Xaa³ is Tyr or Arg,

provided that when Xaa¹ is Tyr and Xaa² is Leu, then Xaa³ is not Arg.

2. The composition according to Claim 1, wherein the composition is used for the treatment of osteoporosis.
3. The pharmaceutical composition according to Claim 1, wherein the peptide is

pyroGlu-His-Trp-Ser-His-Gly-Trp-Tyr-Pro-Gly-NH₂

4. A method of preparing a pharmaceutical composition for the treatment of osteoporosis, which method comprises the mixing of a peptide according to the sequence

pyroGlu-His-Trp-Ser-Xaa¹-Gly-Xaa²-Xaa³-Pro-Gly-NH₂

or a salt thereof

wherein Xaa¹ is His or Tyr,
Xaa² is Trp or Leu, and
Xaa³ is Tyr or Arg,

provided that when Xaa¹ is Tyr and Xaa² is Leu, then Xaa³ is not Arg, with one or more pharmaceutically acceptable excipients.

5. A method of treatment of an individual suffering from osteoporosis, or at risk of so suffering, which comprises the administration to said individual of a therapeutically

effective amount of a pharmaceutical composition which includes a peptide according to the sequence

pyroGlu-His-Trp-Ser-Xaa¹-Gly-Xaa²-Xaa³-Pro-Gly-NH₂

or a salt thereof

wherein Xaa¹ is His or Tyr,
 Xaa² is Trp or Leu, and
 Xaa³ is Tyr or Arg,

provided that when Xaa¹ is Tyr and Xaa² is Leu, then Xaa³ is not Arg, as an active principal.



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